A Simple and Highly Efficient Preparation of Structurally Diverse Aryl β -diketoacids as HIV-1 Integrase Inhibitors[†]

JIANG, Xiao-Hua(姜晓华) LONG, Ya-Qiu*(龙亚秋)

State Key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences, Shanghai 201203, China

In order to provide a facile and practical access to structurally diverse aryl β -diketoacids, An improved and highly efficient oxalylation method was developed which employed commercially available and cheap reagents. The oxalylation of aryl methyl ketones, the key step to construct the pharmacophore of aryl β -diketoacids, was considerably facilitated by a new combination of dimethyl oxalate as an oxalic source and sodium *tert*-butoxide as a base. A wide variety of aryl β -diketoacids bearing different functional groups can be prepared rapidly in high yields at room temperature with this method, which has significant advantages over the previously reported procedures in a wider application range, much less amount of reagents, pretty higher yields and quite shorter reaction time. The bis-aryldiketoacids **3k** and **3l**, readily prepared by this method, displayed interesting and promising inhibitory activities against HIV-1 integrase and HIV-1 replication in cells.

Keywords oxalylation, sodium *tert*-butoxide, dimethyl oxalate, aryl β -diketoacid, HIV-1 integrase inhibitor, bis-diketoacid

Introduction

Aryl diketoacids (ADK) constitute an interesting class of bioactive structural motif. They were initially reported as effective inhibitors of viral polymerases, in particular hepatitis C virus RNA-dependent RNA polymerase (HCV RdRp), hepatitis B virus polymerase (HBV pol) and reverse transcriptase of human immunodeficiency virus (HIV RT).¹ Recently the diketoacids were discovered as the most promising class of HIV-1 integrase inhibitors.^{2,3} HIV-1 integrase (IN) is an essential enzyme for the replication and infection of retroviruses,⁴ so IN is an attractive target for antiviral therapy.⁵ Among the numerous IN inhibitors identified,⁶⁻⁸ the most developed are the aryl diketo (ADK) family (Figure 1),³ among which S-1360 was the first drug as an HIV-1 IN inhibitor to enter clinical trials.^{9,10} These

findings have fuelled the recent surge of interest in the development of HIV-1 IN inhibitors.⁶

The structure-activity relationship study and further development of ADK-based inhibitors require a ready access to structurally diverse aryl β -diketoacids. However, the conventional methods to synthesize ADK compounds, *i.e.* diethyl oxalate or dimethyl oxalate in the presence of NaH or NaOEt or NaOMe,¹¹⁻¹⁵ exhibited very low efficiency for the sterically hindered or halogen-substituted aryl ketones, greatly limiting the preparation of various aryl diketoacids needed for the discovery and development of IN inhibitors. Herein we report our newly developed facile and practical oxalylation method, which employs dimethyl oxalate as a convenient oxalic source and sodium *tert*-butoxide as a base to prepare a wide range of aryl diketoacids in high yields (Scheme 1).



Figure 1 Reported ADK-based HIV IN inhibitors.

* E-mail: yqlong @mail.shcnc.ac.cn; Fax: +86-21-50806876
 Received March 25 2004; revised and accepted July 2, 2004.
 Project supported by Shanghai Municipal Committee of Science and Technology (Nos. 02QB14056 and 03DZ19219), the Chinese Academy of Sciences (KSCX1-SW-11) and the Ministry of Personnel of China.
 [†]Dedicated to Professor Chengye Yuan on the occasion of his 80th birthday.

Scheme 1^a



^{*a*}(i) Procedure A: $(CO_2Me)_2$, NaOCH₃, THF-DME (1 : 1), r.t., \geq 24 h; Procedure B: $(CO_2Me)_2$, NaOBu-*t*, THF-DME (1 : 1), r.t., 10 min—1.5 h (except for **1i**, 12 h); (ii) 1 mol/L NaOH, THF-CH₃OH (1 : 1), r.t., 1 h.

Results and discussion

As part of our program to examine the structural features of the ADK family which are related to IN inhibitory potency and selectivity, we have tried to develop a simple and highly efficient procedure to afford more structurally diverse aryl β -diketoacids conveniently. Recently we reported that tert-butyl methyl oxalate instead of conventional dimethyl oxalate in the presence of sodium methoxide was an efficient system to prepare a variety of aryl diketoacids in good to excellent yields at room temperature.¹⁶ However, this methodology is not effective for electron-withdrawing group substituted acetophenones such as 4-nitroacetophenone and 4-trifluoromethylacetophenone. Furthermore, the key oxalylating agent, *tert*-butyl methyl oxalate needs preparing from oxalyl chloride via two-step reaction. So, it is intriguing to optimize the procedure, starting from commercially available and cheap reagents.

Based on the preliminary study on the mechanism of tert-butyl methyl oxalate mediated oxalylation with sodium methoxide as a base, we proposed that sodium methoxide might react first with the tert-butyl methyl oxalate to generate dimethyl oxalate and sodium *tert*-butoxide in a transesterification, then the resulting sodium tert-butoxide would function as a better base in the reaction (Scheme 2).¹⁶ Enlightened by this idea, we supposed that sodium tert-butoxide could be used directly as a base to promote the dimethyl oxalate mediated coupling reaction. Interestingly, the new combination of dimethyl oxalate and sodium tert-butoxide was found remarkably efficient for the oxalylation of a wide range of aryl methyl ketones, even more efficient than its cognate system of tert-butyl methyl oxalate and sodium methoxide. The positive results of the mechanism-based design further proved our proposed mechanism in turn.

As summarized in Table 1, the new combination of dimethyl oxalate/sodium *tert*-butoxide compares

Scheme 2 Proposed mechanism of the oxalylation mediated by *tert*-butyl methyl oxalate and sodium methoxide



favorably with both the conventional system of dimethyl oxalate/sodium methoxide and our recently reported system of tert-butyl methyl oxalate/sodium methoxide for the preparation of various ADK derivatives. A variety of aryl methyl ketones bearing electron-donating or electron-withdrawing groups as well as sterically crowded and bis-(aryl methyl ketone) examples were screened. The coupling of variously substituted acetophenones with dimethyl oxalate in the presence of sodium tert-butoxide occurred very efficiently at room temperature, using 1 equiv. of aryl ketone, 2 equiv. of dimethyl oxalate, and 2.5 equiv. of sodium tert-butoxide. In the case of using sodium methoxide as a base, the quantity of sodium methoxide was increased to 5 equiv. when dimethyl oxalate was used, whereas the amount of sodium methoxide still required 4 equiv. when tert-butyl methyl oxalate was employed as oxalic source. The optimized mixed solvent of THF and DME (1:1) was used for the standard coupling procedure.

It is clear that, in most cases, shorter time and better yields were achieved for the couplings using sodium tert-butoxide, compared with those using sodium methoxide as a base. Halogen (1a-1c, 1f), electron-withdrawing group (1d-1e), and electron-donating group (1g-1l) substituted acetophenones all exhibited an overwhelming preference for the sodium tert-butoxide/ dimethyl oxalate system. Especially for the preparation of electron-withdrawing group substituted ADK products, the current combination displayed a unique advantage over its cognate system of *tert*-butyl methyl oxalate and sodium methoxide, the former proceeding rapidly in excellent yields, while the latter just affording low to moderate yields after a longer reaction time. The number and the position of the substituents on the phenyl ring did not affect the efficiency. Even the hindered ketones (1a, 1c, 1k—1l) did not attenuate the reactivity. Similar to tert-butyl methyl oxalate and sodium methoxide method, the new oxalylation system of dimethyl oxalate/sodium tert-butoxide showed special advantage in the synthesis of bis-aryl diketoacids (entries 11, 12 in Table 1), which constitute a novel set of ADK-based HIV-1 IN inhibitors.^{14,17} The preparation of bis-diketo acid via dimethyl oxalate in the presence of sodium methoxide failed at room temperature, even after 24 h

980 Chin. J. Chem., 2004, Vol. 22, No. 9

Table 1	Coupling c	of arvl meth	vl ketones w	vith three o	oxalvlation	systems

Entre			Isolated yield (%) (Reaction time)			
Entry	Aryl ketone		\mathbf{I}^{a}	Π^b	III^{c}	- Product
1	F O	1 a	44 (24 h)	95 (10 min)	81 (1.0 h)	2a
2	F	1b	12 (24 h)	100 (20 min)	97 (1.5 h)	2b
3	CIO	1c	0 (24 h)	NA^d	92 (10 min)	2c
4	O ₂ N	1d	30 (24 h)	52 (2 h)	98 (10 min)	2d
5	F ₃ C	1e	18 (24 h)	71 (2 h)	94 (10 min)	2e
6	CI	1f	0 (24 h)	86 (12 h)	93 (0.5 h)	2f
7		1g	33 (24 h)	78 (2 h)	89 (10 min)	2g
8		1h	14 (36 h)	76 (2 h)	82 (10 min) ^e	2h
9	C O H O O	1i	0 (24 h)	92 (3 h) ^f	85 (12 h)	2i
10	H ₃ CO	1j	0 (24 h)	NA	97 (10 min)	2j
11		1k	$0^{g} (24 h)^{h}$	94 (0.5 h) ^h	84 (10 min) ^h	2k



^{*a*} Dimethyl oxalate/sodium methoxide system. The coupling was performed according to procedure A. ^{*b*} *tert*-Butyl methyl oxalate/sodium methoxide system. The data were reported in Ref. 16 and used here as a comparison. ^{*c*} Dimethyl oxalate/sodium *tert*-butoxide system. The coupling was carried out according to procedure B. ^{*d*} Not available. ^{*e*} The isolated yield was obtained with 4 equiv. of NaOMe. ^{*f*} The isolated yield was obtained after 3 h stirring at 40 °C. ^{*g*} NaOMe (10.0 equiv.), dimethyl oxalate (5.0 equiv.), toluene-DME-THF (1 : 1 : 1), 60 °C overnight, the isolated yield 50%. ^{*h*} The coupling of bis-aryl methyl ketones was performed in toluene-DME-THF (1 : 1 : 1) due to solubility, and the quantity of the reagents was adjusted proportionally with the number of ketones.

stirring (1k, 1l). Raising the temperature to 60 $^{\circ}$ C in toluene and DME helped the reaction to proceed in moderate yields (50%—60%) after 12 h heating. However, the coupling mediated by sodium *tert*-butoxide (1k, 1l) resulted in high yields (85%—88%) within a very short time (10 min at room temperature usually).

The final aryl diketoacids can be readily obtained from the hydrolysis of the resulting aryl diketoacid methyl ester with 1 mol/L NaOH in MeOH-THF at room temperature. The typical procedure was examplified by the preparation of aryl diketoacid 3k.

Compared to the recently reported *tert*-butyl methyl oxalate/sodium methoxide method,¹⁶ the dimethyl oxalate/sodium *tert*-butoxide system exhibited an improved efficiency and significant advantages in terms of a wider application range, less amount of reagents and shorter reaction time. Furthermore, the commercially available and cheap reagents employed render this new combination an attractive and practical approach to prepare active ADK-based HIV IN inhibitors in large scale.

Among our synthetic ADK compounds, the bis-aryl diketoacids **3k** and **3l** were chosen to test the HIV-1 IN inhibitory activity. The bioassay was evaluated in an extracellular HIV-1 integrase assay¹⁸ and in a single-cycle replication assay using envelope-deficient HIV-1.¹⁹ The data were summarized in Table 2.²⁰ Both exhibited interesting and promising inhibitory activity against HIV-1 integrase as well as anti-viral efficiency in HIV-infected cells.

Conclusion

In summary, we developed a facile and efficient procedure to prepare variously substituted aryl diketoacids with a new combination of dimethyl oxalate/sodium *tert*-butoxide. The oxalylation was accomplished in high speed with excellent yields under mild conditions. This methodology is applicable to a wide range of aryl methyl ketones bearing electron-donating or electron-withdrawing groups, or bis-(aryl methyl ketone) examples, thus offering a general method for the preparation of structurally diverse ADK-containing derivatives. The bis-aryldiketoacids **3k** and **3l**, readily prepared by this method, displayed interesting and potent inhibitory activity against HIV-1 integrase and antiviral potency in HIV-infected cells. This highly efficient preparation of aryl β -diketoacids using dimethyl oxalate in the presence of sodium *tert*-butoxide will potentially benefit the development of HIV-1 IN inhibitors.

Experimental

¹H NMR spectra were recorded on a Varian 300 MHz or 400 MHz spectrometer. ¹³C NMR spectra were recorded on a Varian Mercury VX 400 MHz spectrometer. Elemental analyses were obtained on a Vario EL analyzer. Melting points (uncorrected) were determined on a Buchi-510 capillary apparatus. IR spectra were recorded on a Bio-Rad FTS-185 spectrophotometer. HRMS (ESI) spectra were obtained on a APEXIII 7.0 TESLA FTMS mass spectrometer. Substituted aryl acetophenones 1a-1f were of commercial grade, and **1g—1** were synthesized according to the literature procedure. The solvents (toluene, THF and DME) were dried over sodium wire and distilled prior to use. Flash column chromatography was performed on silica gel H $(10-40 \mu m)$ with petroleum ether-ethyl acetate system as eluent.

General procedure for oxalylation of aryl methyl ketone

Procedure A: To a stirred solution of NaOCH₃ (5.0 equiv.) in anhydrous THF at room temperature was added dropwise the mixture of dimethyl oxalate (2.0 equiv.) and aryl methyl ketone (1.0 equiv.) in DME. The resulting orange-yellow mixture was stirred at room temperature for 24 h at least. The reaction was quenched with 1.0 mol/L aqueous HCl solution and extracted with CH₂Cl₂. The combined organic layers were washed with saturated aqueous NaHCO₃ solution and brine

Table 2 Inhibition of HIV-1 integrase catalytic activities and HIV-1 replication in cells by the synthesized bis-aryidiketoa

Compd –	Inhibition of integrase catalytic activities ^a			Antiviral activities ^b		
	3'-Processing $IC_{50}/(\mu mol \cdot L^{-1})^{c}$	Strand transfer $IC_{50}/(\mu mol \bullet L^{-1})^c$	SI^d	$EC_{50}/(\mu mol \bullet L^{-1})^e$	$CC_{50}/(\mu mol \bullet L^{-1})^f$	
3k	1.8 ± 0.9	0.3 ± 0.2	6	39 ± 3	>200	
31	7.3 ± 2.3	0.4 ± 0.1	18	>200	>200	

^{*a*} Assays were performed with recombinant HIV-1 integrase (0.2 μ mol·L⁻¹) preincubated with the inhibitor in reaction buffer at 30 °C for 30 min. Then, 20 nmol·L⁻¹ of the 5'-end ³²P-labeled linear oligonucleotide substrate was added, and incubation was continued for an additional 1 h. For details see Ref. 19. ^{*b*} The anti-HIV activity was evaluated in human T cell line CEM-SS infected with HIV-1 as described by Weislow *et al.*^{20 c} IC₅₀: Inhibitory concentration 50% (inhibition of purified integrase). ^{*d*} SI: Sensitivity index=IC₅₀ 3'-P/IC₅₀ ST. ^{*e*} EC₅₀: Effective concentration 50% (protection of HIV-1 infected CEM cells). ^{*f*} CC₅₀: Cytotoxic concentration 50% (toxicity to uninfected CEM cells).

respectively, dried over Na₂SO₄ and concentrated under vacuum. Purification by silica gel flash chromatography provided the desired product.

Procedure B: To a stirred mixture of *tert*-BuONa (2.5 equiv.) and dimethyl oxalate (2.0 equiv.) in anhydrous THF at 0 °C was added dropwise aryl methyl ketone (1.0 equiv.) in DME. The resulting orange-yellow mixture was stirred at room temperature for 1.5 h at most (for 1i, 12 h). The reaction was quenched with 1.0 mol/L aqueous HCl solution and extracted with CH₂Cl₂. The combined organic layers were washed with saturated aqueous NaHCO₃ solution and brine respectively, dried over Na₂SO₄ and concentrated under vacuum. Purification by silica gel flash chromatography provided the desired product.

4-(2-Chlorophenyl)-2-hydroxy-4-oxo-but-2-enoic acid methyl ester (2c): White solid. m.p. 80—81 °C. ¹H NMR (400 MHz, CDCl₃) δ: 3.93 (s, 3H), 6.97 (s, 1H), 7.35—7.39 (m, 1H), 7.42—7.49 (m, 2H), 7.63— 7.65 (m, 1H), 14.60 (br, 1H); ¹³C NMR (100 MHz, CDCl₃) δ: 192.54, 166.99, 162.07, 135.52, 132.62, 131.94, 130.79, 130.05, 126.93, 103.05, 53.32; IR (KBr) v: 3444, 2964, 1726, 1591, 1432, 1274, 1043, 733 cm⁻¹; EI-MS m/z (%): 240 (M⁺, 0.59), 205 (25.88), 181 (100.0), 139 (30.12); Anal. calcd for C₁₁H₉ClO₄: C 54.90, H 3.77; found C 55.12, H 3.88.

4-(3-Benzyloxy-4-methoxy-phenyl)-2-hydroxy-4oxo-but-2-enoic acid methyl ester (2j): Yellow solid. m.p. 149—150 °C. ¹H NMR (400 MHz, CDCl₃) & 3.93 (s, 3H), 3.96 (s, 3H), 5.25 (s, 2H), 6.94 (d, J=8.7 Hz, 1H), 7.03 (s, 1H), 7.30—7.35 (m, 1H), 7.36—7.45 (m, 4H), 7.56 (s, 1H), 7.58 (d, J=2.1 Hz, 1H), 14.65 (br, 1H); ¹³C NMR (100 MHz, CDCl₃) & 190.36, 166.49, 162.62, 153.04, 149.51, 135.76, 128.51 (×2), 128.02, 127.02 (×2), 122.52, 112.29, 110.18, 98.03, 70.83, 56.13, 53.16; IR (KBr) *v*: 3466, 2947, 1747, 1595, 1514, 1427, 1259, 1003, 771, 748 cm⁻¹; EI-MS *m/z* (%): 342 (M⁺, 19.95); 283 (6.05); 91 (100); Anal. calcd for C₁₉H₁₈O₆: C 66.66, H 5.30; found C 66.71, H 5.02.

2-Hydroxy-4-(3-{3-[3-(3-hydroxy-3-methoxycarbonylacryloyl)phenoxymethyl]benzyloxy}phenyl)-4-oxo-but-2-enoic acid methyl ester (2l): Pale yellow solid. m.p. 145—148 °C. ¹H NMR (400 MHz, CDCl₃) δ: 3.95 (s, 6H), 5.16 (s, 4H), 7.06 (s, 2H), 7.21—7.24 (m, 2H), 7.39—7.45 (m, 5H), 7.56 (s, 1H), 7.58—7.61 (m, 4H), 15.20 (br, 2H); ¹³C NMR (100 MHz, CDCl₃) δ: 190.32 (×2), 168.55 (×2), 162.32 (×2), 158.75 (×2), 136.69 (×2), 136.10 (×2), 129.79 (×2), 128.87 (×2), 127.11 (×2), 126.33 (×2), 120.70, 120.62, 113.26 (×2), 98.23 (×2), 69.97 (×2), 53.26 (×2); IR (KBr) *v* : 951, 1745, 1605, 1450, 1271, 1211, 1059, 885, 771 cm⁻¹; EI-MS *m*/*z* (%): 531 (0.1), 374 (14.0), 239 (100.0), 221 (12.0), 163 (28.0), 121 (10.0), 104 (36.0); Anal. calcd for C₃₀H₂₆O₁₀: C 65.93, H 4.80; found C 65.76, H 4.78.

2-Hydroxy-4-(3-{2-[3-(3-carboxy-3-hydroxy-bonylacryloyl)phenoxymethyl]benzyloxy}phenyl)-4oxo-but-2-enoic acid (3k): A solution of compound 2k: (0.164 g, 0.30 mmol) in THF/CH₃OH (1:1) (4.0 mL) was treated with 1.0 mol/L NaOH (3.0 mL, 3.0 mmol). The reaction mixture was stirred for 1 h at room temperature, then extracted with ether. The water phase was acidified with 2.0 mol/L HCl solution to pH=1-2 and extracted with ethyl acetate. The combined organic layers were washed with brine and dried over Na₂SO₄ and the solvent was removed under vacuum. The residue was recrystallized from petroleum ether/methane dichloride to give compound 3k as a pale yellow solid (0.109 g, yield 70.0%). m.p. 186–188 °C. ¹H NMR (400 MHz, CDCl₃) δ: 5.36 (s, 4H), 7.06 (s, 2H), 7.32-7.35 (m, 2H), 7.39-7.41 (q, J=13.40 Hz, 2H), 7.45-7.49 (m, 2H), 7.55-7.59 (m, 3H), 7.61-7.64 (m, 3H); ¹³C NMR (100 MHz, CDCl₃+DMSO- d_6) δ : 190.35 (× 2), 169.47 (×2), 163.22 (×2), 158.44 (×2), 136.04 (× 2), 134.29 (×2), 129.63 (×2), 128.88 (×2), 128.30 (× 2), 120.31 (×2), 112.98 (×2), 97.78 (×2), 67.82 (×2); IR (KBr) v: 3539, 3466, 1701, 1624, 1574, 1290, 1261, 899, 775 cm⁻¹, HR-ESI-MS calcd for $C_{28}H_{22}O_{10}$ $(M+Na)^+$ 541.1111, found 541.1113.

2-Hydroxy-4-(3-{3-[3-(3-carboxy-3-hydroxy-bonylacryloyl)phenoxymethyl]benzyloxy}phenyl)-4oxo-but-2-enoic acid (3l): Compound 3l was obtained from the hydrolysis of compound 2l with 1.0 mol/L aqueous NaOH solution (3.2 mL, 3.2 mmol) in THF/CH₃OH (1 : 1) (4.0 mL) analogously to compound 3k. Recrystallization from petroleum ether/methane dichloride afforded compound **3l** as a pale yellow solid (0.122 g, yield 73.5%). m.p. 188—191 °C. ¹H NMR (400 MHz, DMSO- d_6) & 5.23 (s, 4H), 7.09 (s, 2H), 7.33 —7.36 (m, 2H), 7.48—7.51 (m, 6H), 7.63—7.68 (m, 4H); ¹³C NMR (100 MHz, DMSO- d_6) & 190.33 (×2), 163.51 (×2), 158.65 (×4), 136.50 (×4), 130.27 (×2), 127.93 (×6), 120.42 (×2), 113.12 (×2), 98.28 (×2) 69.21 (×2); IR (KBr) v: 3533, 2895, 2459, 1685, 1624, 1574, 1286, 1263, 901, 775 cm⁻¹; HR-ESI-MS calcd for C₂₈H₂₂O₁₀ (M+Na)⁺ 541.1111, found 541.1108.

References

- Altamura, S.; Tomei, L.; Koch, U.; Neuner, P. J. S.; Summa,
 V. WO 0006529 A1, 2000 [Chem. Abstr. 2000, 132, 132323].
- 2 Goldgur, Y.; Craigie, R.; Cohen, G. H.; Fujiwara, T.; Yoshinaga, T.; Fujishita, T.; Sugimoto, H.; Endo, T.; Murai, H.; Davies, D. R. *Proc. Natl. Acad. Sci. U.S.A.* **1999**, *96*, 13040.
- 3 Hazuda, D. J.; Felock, P.; Witmer, M.; Wolfe, A.; Stillmock, K.; Grobler, J. A.; Espeseth, A.; Gabryelski, L.; Schleif, W.; Blau, C.; Miller, M. D. *Science (Washington, D. C.)*, 2000, 287, 646.
- 4 Esposito, D.; Craigie, R. Adv. Virus Res. 1999, 52, 319.
- 5 Young, S. D. Curr. Opin. Drug Discov. Devel. 2001, 4, 402.
- 6 Neamati, N. Expert Opin. Invest. Drugs 2002, 12, 709.
- 7 Neamati, N. Expert Opin. Invest. Drugs 2001, 10, 281.
- 8 Dayam, R.; Neamati, N. Curr. Pharm. Design 2003, 9, 1789.
- 9 Yoshinaga, T.; Sato, A.; Fujishita, T.; Fujiwara, T. In Vitro Activity of A New HIV-1 Integrase Inhibitor in Clinical Development, 9th Conference on Retroviruses and Opportunistic Infections, Seattle, USA, 2002.
- 10 Billich, A. Curr. Opin. Invest. Drugs (Thomson Current

Drugs), 2003, 4, 206.

- Williams, H. W. R.; Eichler, E.; Randall, W. C.; Rooney, C. S.; Cragoe, E. J., Jr.; Streeter, K. B.; Schwam, H.; Michelson, S. R.; Patchett, A. A.; Taub, D. J. Med. Chem. 1983, 26, 1196.
- 12 Wai, J. S.; Egbertson, M. S.; Payne, L. S.; Fisher, T. E.; Embrey, M. W.; Tran, L. O.; Melamed, J. Y.; Langford, H. M.; Guare, J. P., Jr.; Zhuang, L.; Grey, V. E.; Vacca, J. P.; Holloway, M. K.; Naylor-Olsen, A. M.; Hazuda, D. J.; Felock, P. J.; Wolfe, A. L.; Stillmock, K. A.; Schleif, W. A.; Gabryelski, L. J.; Young, S. D. J. Med. Chem. 2000, 43, 4923.
- Fujishita, T.; Yoshinaga, T.; Yamauchi, H. WO 9950245 A1,
 1999 [Chem. Abstr. 1999, 131, 271806].
- 14 Pais, G. C. G.; Zhang, X.; Marchand, C.; Neamati, N.; Cowansage, K.; Svarovskaia, E. S.; Pathak, V. K.; Tang, Y.; Nicklaus, M.; Pommier, Y.; Burke, T. R., Jr. *J. Med. Chem.* 2002, 45, 3184.
- 15 Yuan, J.; Gulianello, M.; De Lombaert, S.; Brodbeck, R.; Kieltyka, A.; Hodgetts, K. J. *Bioorg. Med. Chem. Lett.* 2002, 12, 2133.
- 16 Jiang, X.-H.; Song, L.-D.; Long, Y.-Q. J. Org. Chem. 2003, 68, 7555.
- Marchand, C.; Zhang, X.; Pais, G. C. G.; Cowansage, K.; Neamati, N.; Burke, T. R., Jr.; Pommier, Y. J. Biol. Chem. 2002, 277, 12596.
- Marchand, C.; Neamati, N.; Pommier, Y. *Methods Enzymol.* 2001, *340*, 624.
- Weislow, O. S.; Kiser, R.; Fine, D. L.; Bader, J.; Shoemaker, R. H.; Boyd, M. R. *J. Natl. Cancer Inst.* **1989**, *81*, 577.
- 20 Long, Y.-Q.; Jiang, X.-H.; Dayam, R.; Sanchez, T.; Shoemaker, R.; Sei, S.; Neamati, N. J. Med. Chem. 2004, 67, 2561.

(E0403251 LI, W. H.)